
Microfluidic image cytometry for quantitative single-cell profiling of human pluripotent stem cells in chemically defined conditions.

Journal: Lab Chip

Publication Year: 2010

Authors: Ken-ichiro Kamei, Minori Ohashi, Eric Gschweng, Quinn Ho, Jane Suh, Jinghua Tang, Zeta Tak For Yu, Amander T Clark, April D Pyle, Michael A Teitell, Ki-Bum Lee, Owen N Witte, Hsian-Rong Tseng

PubMed link: 20390128

Funding Grants: Role of Mitochondria in Self-Renewal Versus Differentiation of Human Embryonic Stem Cells, Generation of Pluripotent Cell Lines from Human Embryos, Microfluidic Platform for Screening Chemically Defined Conditions that Facilitate Clonal Expansion of Human Pluripotent Stem Cells, Mitochondrial Metabolism in hESC and hiPSC Differentiation, Reprogramming, and Cancer

Public Summary:

Scientific Abstract:

Microfluidic image cytometry (MIC) has been developed to study phenotypes of various hPSC lines by screening several chemically defined serum/feeder-free conditions. A chemically defined hPSC culture was established using 20 ng mL⁻¹ of bFGF on 20 microg mL⁻¹ of Matrigel to grow hPSCs over a week in an undifferentiated state. Following hPSC culture, we conducted quantitative MIC to perform a single cell profiling of simultaneously detected protein expression (OCT4 and SSEA1). Using clustering analysis, we were able to systematically compare the characteristics of various hPSC lines in different conditions.

Source URL: <https://www.cirm.ca.gov/about-cirm/publications/microfluidic-image-cytometry-quantitative-single-cell-profiling-human>